

DNA SEQUENCE SUBMISSION FORM – INTERNAL

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NAME: _____ DATE: _____

PI/SUPERVISOR: _____ TELEPHONE #: _____

E-MAIL: _____ ROOM #: _____

ACCOUNT NUMBER: **(please fill out the entire MOSAIC CHARTFIELD STRING)**

Fund(2)	Account(6)	Department(5)	Project(8) or Program(5)	PC Bus.Unit(5) (for Projects only)
Example: 85	600001	10999	20099999	RFHSC

EDITING REQUEST** WITH (\$15.00) WITHOUT (\$7.50)
 RESULTS E-MAILED TEXT FILE ABI FILE
 PRINTOUTS YES

SPECIAL CONSIDERATION: 1) higher than 65% GC content (add DMSO) YES
 2) Secondary Structure (hairpin protocol) YES (\$9.00 per rxn, 1 to 2 additional days)

#	Name of DNA (max 8 characters)	Name of PRIMER (max 8 characters)	#	Name of DNA (max 8 characters)	Name of PRIMER (max 8 characters)
1			9		
2			10		
3			11		
4			12		
5			13		
6			14		
7			15		
8			16		

DNA & PRIMER - should be suspended in Water or 0.1x Elution buffer. We require **5µl** of DNA and **5µl primer per reaction**.

Primer concentration: 1 picomole/µl = [1 µM]

DNA concentration: 1ng/µl per 100 bases

DNA Concentration - because sequencing is unidirectional the **number of copies of your template** in the sample becomes very important, too little DNA will result in no-signal. Too much DNA can result in a short read or a smear or no signal. Note that the input concentration should be based on the TOTAL size of the template DNA in the sample; e.g. give us 5µl @ 30 ng/µl if your plasmid + insert is 3 kb. **UV absorbance-based quantification is not always accurate!** Better methods are Qubit Fluorometric or gel-based quantification.

Please indicate your TOTAL template size

ds DNA approx. size _____

PCR products approx. size _____

**Editing is the correction of miscalled bases. These problems generally occur at the beginning and end of the sequence but can occur anywhere.

Note - you require the ABI file and the correct computer software to do your own editing.

SAMPLE STORAGE: we keep the DNA samples and primers for one month after they have been processed.